LIVESTOCK SUSTAINABILITY: HAEMOPARASITIC STATUS OF CATTLE IN ABATTOIR

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Abstract-Beef accounts for nearly half of all meat consumed globally, so cattle play an important role in the meat supply and livestock industry. The reduction in cattle productivity is of a global concern. Therefore, the study aimed to investigate the haemoparasitic statusof cattle in the abattoir of Adodo Ota, Ogun State Nigeria in other to assess and promote food and livestock sustainability.Sixty (60) blood samples were taken at random from the slaughtered and jugular veins of 60 cattle. Thin film was made, air dried then fixed in 100% methanol for 60 seconds, and stained with quality grade 10% Giemsa stain, slides were viewed under ×40 and ×100 power of a binocular microscope atOmega Medical Diagnostics Laboratory. Heamoparasites intensity in cattle indicates 25% (15) infection with E.wenyoni, 11.7% (07) infection with H.bovis, 16.7% (10)infection with mixed parasites of Eperythrozoon and Haemobartonella while8.3% (05) accounting for the lowest percentage infection with T.congolense. High haemoparasitic prevalence was seen in Cows 70% than Bull 57.5%. Heamoparaitic status of cattle from the study signifies that haemoparasites remain one the biggest threat to food and livestock sustainability, and a continuous source of veterinary and public health concern.

Keywords-Abattoir, cattle, haemoparasite, infection, parasite intensity, prevalence

I. INTRODUCTION

Cattle are domestic ungulates mammals, a member of the family Bovidae, they are large grass-eating mammals with two-toed or cloven hooves and a four-chambered stomach which are an adaptation to support the digestion of plant materials [1]. Cattle are raised as livestock globally for various purposes which include the source of animal protein, dairy product, and income [2]. Their byproducts such as hoof, bones, blood, hides, and skin are also widely used as raw materials [3].

According to Ugochukwu&Sidney [3],cattle meat widely referred to as Beef is considered to be the third most widely consumed livestock meat globally, accounting for approximately25% of meat production, meat is rich in total protein and essential body minerals (zinc, iron, vitamin B complex, and phosphorus) among others. Hamoparaites(Blood parasites) are one of the major parasites posing substantial economic concern and impact on livestock management, they are estimated to affect 80% of the cattle population globally [4,5].

Haemoparasites are a diverse community of species characterized by the presence of single or multiple developmental stages in the bloodstream of animals and humans[6].They cause economic losses as a result of an elevatedrate of morbidity and mortality;incurred losses have led to the attenuation in the further development of the livestock sector in most countries [7],they are also responsible for immunosuppression, retarded growth, low milk production and weight loss[8].

Haemoparasites affecting livestock has been documented by various researchers [2,6,9-11], some of which includes.*Aegyptianella species*, *Anaplasma species*, *Babesia species*, *Ehrlichia species*, *Haemobartonella species*, *Haemoproteus species*, *Hepatozoon species*, *Leishmania species*, *Leucocytozoon species*, *Plasmodium species*,*Schistosoma species*, *Theileria species*, *andTrypanosoma species*.

Hamoparasite in cattle has also been recently reportedin northern Nigeria, Kamaniet al.[12]in another study reported 25.7% prevalence rate of infection amongstudied cattle in North-Central, Nigeria. Ola-Fadunsinet al. [6], reported 26.95% prevalence rate of haemoparasitic infections n cattle from a study in Plateau Nigeria.Ugochukwu&Sidney [3].reported a State. prevalence rate of 6.68% in cattle from the abattoir of Oyo state, South west Nigeria. Sam-Woboet al.[2]reported 27.8%haemoprasite prevalence in cattle abattoir of Abeokuta, Southwest from the Nigeria.Although haemoparasitestudies in cattle have been reported in various parts of Nigeria, the continuous need for food and livestock sustainability, and qualitative

livestock production is imperative.Hence the purpose of the study

II. METHODOLOGY

A. Study Area

The study was carried out in an abattoir of Adodo-Ota Local Government Area (LGA)of Ogun State, Southwestern Nigeria. This LGA is situated in the tropical region, between the equator and the Greenwich Meridian at 60 47N and 20 53E and 30 18E, respectively. It has a total area of 1,010.4 square kilometres of flat land and approximately 252.6 square kilometres of poor terrain, with 10% riverine and 4% hilly regions. The abattoir was chosen for cattle blood sample collection because of the high constant slaughtering activities. The study was carried out in February 2020.

B. Blood Sample Collection and Haemoparasite Examination

Blood samples were collected randomly from both slaughtered and the jugular veins ofsixty (60) cattle sold in the abattoir of Adodo-Ota, Ogun State Nigeria. Samples from slaughtered cattle were obtained directly at the point of slaughter using a sterile EDTA bottle after allowing a small amount of blood flow in order to prevent contamination. A sterile syringe and needle were used to collect samples from the jugular veins, which were then placed in a sterile EDTA container. The blood samples were transported to Omega Medical Diagnostics Laboratory for examination. Thin film was made, air dried then fixed in 100% methanol for 60 seconds, and stained with quality grade 10% Giemsa stain as described by Sam-Wobo et al.[2], slides were viewed under ×40 and ×100 objective lens of a binocular microscope (Olympus Corporation, Hamburg, Germany).

C. Data management and Analysis

To produce frequency and percentages, the obtained data were analyzed using the Statistical Package for Social Sciences version 21 software. Chi-Squared test was used to determine the prevalence of the condition between genders and across ages.

III. RESULTS AND DISCUSSION

A. Statistic of Cattle Used for the Study

The statistics of cattle used for the studies are presented in table 1.The genderrevealed 40 (66.7%) of the cattle were Bulls while 20 (33.3%) were Cows. Age statistics revealed 12 (20%) an equal highest percentage to be 2 and 4 years of age respectively, 14 (23.3%) were 3 years, 15 (25%) were 5 years of age, 3 (5%) were 6 years of age while 4 (6.67%) which is the lowest age percentage were 7.

 TABLE 1

 STATISTICS OF CATTLE USED FOR THE STUDY

B. Haemoparasitic Intensity in Cattle

Gender Frequency **Cumulative %** Percentage 40 Bull (M) 66.7 66.7 20 33.3 100.0 Cow (F) Total 60 100 Age (years) 12 20.0 20.0 Two Three 14 23.3 43.3 Four 12 20.0 63.3 Five 15 25.0 88.3 03 5.0 93.3 Six 04 6.67 100.0 Seven 100 60 Total

The intensity of Haemoparasite in cattle presented in Table 2. revealed 37 (61.7 %) of the cattle were being infected. 15 (25 %) of which were infected with Eperythrozoon wenyoni indicating the highest percentage, 07 (11.7%)were infected withHaemobartonellabovi, 10 (16.7%)were infected with a mixed parasite of Eperythrozoon and Haemobartonella while 05 (8.3%) which is the lowest percentage of cattle infected with T. congolense.

TABLE 2HAEMOPARASITE INTENSITY IN CATTLE

Haemoparasite	Frequency	Percentage	Cumulative %
E. wenyoni	15	25.0	25.0
H. bovi	07	11.7	36.7
Eperythrozoon&Ha emobartonella	10	16.7	53.3
T. congolense	05	8.3	61.7
Total	37	61.7	

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C. HaemoparasiticPrevalence in Cattle

Haemoparasitic prevalence between gender and across ages of cattle are presented in Tables 3 and 4, shows that E. wenyoni(27.5%, 20%) and T. congolenses(10%, 5%) were prevalent in bull (m) than the cow respectively while Н. bovi(7.5%, 20%)andEperythrozoon&Haemobartonella (12.5, 25%) were less prevalent in bull than the cow respectively. The overall haemoparasitic prevalence shows that the cow, 14 (70%) were more prevalent than the bull 23 (57.5%). Haemoparasiticprevalence across ages of cattle indicate that E. wenyonihas the highest prevalence in age 6 (66.7 %) followed by age 3 (28.6%) and equal in age 2 and 4 at 25% respectively, while the lowest haemoparasitic prevalence was recorded in age 5 (20%). H. bovihas the highest prevalence in age 3 (28.5%) followed by age 4(16.7%) while the lowest was found in age 5(6.7%). Eperythrozoon & Haemobartonella prevalence was highly recorded in age 5(40%) followed by age 3 (14.3%) while the lowest with an equal prevalence was recorded in both age 2 (8.3%) and age 4 (8.3 %). Finally, T. congolense prevalence was higher in cattle age 4 (25%)and lesser in age 2 (16.7%). Although there was difference in haemoparasitic prevalence between gender, and also across ages but statistically not significant at p<0.05.

D. Discussions

The findings presented in the study show that the majority of the cattle population for the study were Bulls. The highest percentages of the cattle were five years of age followed by three years. Ages two, four, six, and seven years accounted for the lowest percentage of the cattle population. This is not in agreement with the work of El-Metenawy[13]whosehighest age percentage was one and two.

Haemoparasitessuch as *Haemobartonella*, *Eperythrozoon*, and *Trypanosoma*species, which are extracellular parasites of erythrocytes pose detrimental effects on susceptible hosts, this varies from mild effect to death [14]. From the study, 38.3 % of the cattle showed no haemoparasiticinfection, while 61.7 % of the cattle were infected. The report was higher than the report made by Kamani*et al.*[12]in Northern Nigeria (25.7%),

Ugochukwu&Sidney [3]in Oyo state (6.68%), Sam-Woboet al.[2]inAbeokuta, South western Nigeria (27.8%) and Ola-Fadunsinet al. [6] in Plateau State, North Central, Nigeria. The highest percentage of the cattle were infected with *E. wenyoni*. This is in line with the work of Al-khalifaet al. [15]whoalso reported 1-4% infectionwith *E. wenyoni*inan examined cattle in Saudi Arabia. Similarly,Hasan [8]reported 28.3% of infected cattlewith *E. wenyoni* inMosul, Iraq.

16.7% of cattle from the study were observed to be infected with mixed haemoparasite of *Eperythrozoon* and *Haemobartonella*. This is similar to the work of Hasan [8]in Mosul, Iraq. *Eperythrozoon* and *Haemobartonella* observed in the study is traceable to environmental suitability enhancing breeding and survivalof vectors (mosquitoes and tick). According to Kahn & Line [16], ticks (*Rhipicephalus*) are vectors of *Eperythrozoon* for cattle, Transmission may also occur through blood contaminated instruments during vaccination.

Haemobartonellaboviis a rod chain in appearance and is usually seenaround the peripheral of Erythrocytes. In the study, the intensity of infection with *H.bovis* in cattle was 11.7% which is higher than the report made byHasan [8] who reported 1.81% of *H.bovi*infection in cattle from Iraq.

Trypanosomacongolenseis described as the smallest of the African Trypanosomes ranging from approximately 9-18 µ in length. It is considered the most economically important animal Trypanosomiasis[17]. According to Rodistitiset al. [18], infecting cattle with trypanosomes is possible even in the absence of Tsetse fly (Glossina), biting flies such as Tabanidas, Stomoxyinae and Hipoboscidae are mechanical vectors transmitting trypanosomes. T. congolense is responsible for the least amount of infection in the cattle. They appear in peripheral smears in sizes ranging from 8 to 10μ with the absence of a free flagellum as defined by Soulsby [19]. This is in agreement with the work of Ohaeri [20] who recorded the occurrence of T. congolense infection in cattlein Abia State, Nigeria. But not in agreement with the high report of 50% T. congolenseinfection in cattle from the work of Samdiet al. [21]in Kaduna, Nigeria.

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The overallhaemoparasitic prevalence in cattle from the study was higher in cows (70%) thanbull (57.5%).Highhaemoparasitic prevalence in cows could betraceable to the extensive breeding behaviour adopted for economic purposes (calving and production of milk). The susceptibility of cows might also be attributed to altered immunity from pregnancy and lactation stress [22].This is in good tandemwith the report of Okorafor&Nzeako [22] who recorded greater parasite prevalence in cows (4.44%) than bulls (2.22%) in Oyo State, Nigeria.

The overall haemoparasitic prevalence across ages of cattle in the study was high in the cattle of age 3 and 4 years accounting for 71.5% and 75%. This is in contrast to the work of Geoffrey *et al.*[23]with a record of slightly higher parasite prevalence in cattle less than 2 years tothose above in Uganda. It also confirms another blood parasite study in Uganda by Keneth*et al.* [7] who documented a prevalence of 60.4% in adult cattle greater than 1.5 years over 4% in juvenile cattle less than 1.5 years.

IV. CONCLUSIONS

Haemoparasitic diseases constitute a disease entity of considerable economic importance globally and this accounts for a major mitigating factor in maintaining exotic cattle production which in turn affect production of beef, dairies etc. Therefore, there is a pressing need to develop and introduce a successful intervention in cattle health management to enhance production.

Reference

- Adewole SO, Fabumuyi AO, Agunbiade RO, Ayeni SK. Occurrence of Rhabditisdubia in Cattle Dungs in Lagos State, Nigeria. J Env Earth Sci 2014; 4: 8-9
- Sam-Wobo SO, Uyigue J, Surakat OA, Adekunle NO, Mogaji HO. Babesiosis and other Heamoparasitic Disease in a Cattle Slaughtering Abattoir in Abeokuta, Nigeria. Int. J. Trop. Disease Health 2016; 18(2): 1-5.
- Ugochukwu PO, Sidney, ON. Prevalence ofHaemoparasitesof Cattle from Three Abattoirs in Ibadan Metropolis, Oyo State, Nigeria. Int J Sci. Res. Env. Sci. 2014; 2: 244-249
- Kasozi KI, Matovu E, Tayebwa DS, Natuhwera J, Mugezi I, Mahero M. Epidemiology of Increasing Hemo-Parasite Burden in Ugandan Cattle. Open J Vet. Med. 2014; 4: 220-231.

- Muhanguzi D, Picozzi K, Hatendorf J, Thrusfield M, Welburn SC, Kabasa JD, Waiswa C. Prevalence and Spatial Distribution of Theileriaparvain Cattle under Crop-Livestock Farming systems in Tororo District, Eastern Uganda. Parasit. Vect. 2014;7: 91.
- Ola-Fadunsin SD, Karaye PG, Dogo GA. Haemoparasite Fauna of Domestic Animals in Plateau State, North Central Nigeria. B. J. Pure and Applied Sci. 2018; 11 (2): 156-161.
- Keneth IK, Enoch M, Dickson ST, Jemimah N, Israel M, Michael M. Epidemiology of Increasing Hemo-Parasite Burden in Ugandan Cattle, Open J Vet. Med. 2014; 4: 220-231
- Hasan MH. Diagnosis of some blood parasites in cattle and sheep in Mosul, Iraq J Vet Sci. 2012; 26: 57-61
- Pam VA, Ogbu KI, Igeh CP, Bot CJ, Vincent G. Prevalence of Endoparasites of Horses in Jos North and Jos South Local Government Areas of Plateau State Nigeria. J Anim. Sci. Adv.2013; 3(2): 103-108
- Bhattacharjee K, Sarmah PC. Prevalence of haemoparasites in pet, working and stray dogs of Assam and North-East India: A hospital based study. Vet. World 2013; 6(11): 874-878.
- Opara MN, Santali A, Mohammed BR, Jegede OC. Prevalence of Haemoparasites of Small Ruminants in LafiaNassarawa State: A Guinea Savannah Zone of Nigeria. J Vet. Adv. 2016; 6(6); 1251-1257
- 12. Kamani J, Sannusi A, Egwu OK, Dogo GI, Tanko T J, Kemza S, Tafarki AE, Gbise DS. Prevalence and Significance of Haemoparasitic Infections of Cattle in North- Central, Nigeria. Vet. World 2010; 3(10): 445-448.
- El-Metenawy TM. Prevalence of blood parasites among cattle at the central area of Saudi Arabia. Vet. Parasit. 2000; 87: 231–236
- 14. Robson S, Kemp B. Eperythrozoonsis in sheep. State of New South Wales:
 2007;
 http://www.dpi.nsw-gov.au/primefacts(Accessed Mar. 2021).
- Al-Khalifa MS, Hussein HS, Diab FM, Khalil GM. Blood parasites of livestock in certain regions in Saudi Arabia Saudi. Saudi J Bio. Sci 2009; 16: 63-67
- Khan CM, Line S. The Merck Veterinary Manual, 10th ed. Publisher: Merck & Co., Inc. Whitehouse Station, N.J. 2010. pp. 2945
- Gillingwater K, Mamabolo MV, Majiwa PA. Prevalence of mixed Trypanosomacongolenseinfections in livestock and tsetse in kwazulu-Natal, South Africa. J South Afr. Vet. Assoc. 2010; 81(4): 219-23.
- Rodistitis OM, Gay CC, Hinchcliff KW, Constabl PD. Veterinary Medicine A text book of the diseases of cattle, horses, sheep, pigs and goats. 10th ed. Philadephia, WB, Saunders company; 2007. pp. 1532-1533
- Soulsby FJL, Helminths, Arthropods and Protozoa of domesticated animals. 7th ed. London, Philadelphia, Toronto. 1982. pp 754-756
- Ohaeri CC. Prevalence of trypanosomiasis in Ruminants in parts of Abia state, Nigeria. J Anim. Vet. 2010; 9 (18): 2422-2426
- Samdi SM, Fajinmi, AO, Kalejaye JO, Wayo B, Haruna MK, Yarnap JE, Mshelia WP, Usman AO, Harma SM, Jijitar A,

https://doi-ds.org/doilink/07.2021-85599365/IJMRE

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Oqunwole R, Ovbagbedia RP, Bizi R. Prevalence of trypanosomosis in cattle at slaughter in kaduna central Abattoir. Asian J Ani. Sci. 2011; (5): 162-165

- Okorafor UP, Nzeako SO. Prevalence of Haemoparasites of Cattle from Three Abattoirs in Ibadan Metropolis, Oyo State, Nigeria. Int. J Sci Res. Env. Sci. 2014; 2(7): 244-249
- 23. Geoffrey W, James O, Samuel GO, Gabriel T, Susan N, Salvatory A, Tony LG. Prevalence and Risk Factors Associated with Hemoparasites in Cattle and Goats at the edge of Kibale National Park, Western Uganda. J Parasit. 2017; 103(1): 69–74

TABLE 3

Haemoparasite		Bull	Cow	Total	
	No. Examined	40	20	60	
E. wenyoni	No. Positive	11	04	15	
	% of infection	27.5	20	25	
H. bovi	No. Positive	03	04	07	
	% of infection	7.5	20	11.7	
Eperythrozoon&Ha	No. Positive	05	05	10	
emobartonella	% of infection	12.5	25	16.7	
T. congolenses	No. Positive	04	01	05	
	% of infection	10	05	8.3	
	Total infection (%)	23 (57.5)	14 (70)	37(61.7	

HAEMOPARASITIC PREVALENCE BETWEEN GENDERS OF CATTLE

HAEMOPARASITIC PREVALENCE ACROSS AGESOF CATTLE

	Age (years)						
	Two	Three	Four	Five	Six	Seven	Total
No. Examined	12	14	12	15	03	04	60
No. Positive	03	04	03	03	02	-	15
% of infection	25	28.6	25	20	66.7	-	25
No. Positive	-	04	02	01	-	-	07
% of infection	-	28.6	16.7	6.7	-	-	11.7
No. Positive	01	02	01	06	-	-	10
% of infection	8.3	14.3	8.3	40	-	-	16.7
No. Positive	02	-	03	-	-	-	05
% of infection	16.7	-	25	-	-	-	8.3
Total infection (%)	06 (50)	10 (71.5)	9 (75)	10(66.7)	02(66.7)	-	37(61.7)
-	No. Positive % of infection No. Positive % of infection No. Positive % of infection No. Positive	No. Examined12No. Positive03% of infection25No. Positive-% of infection-No. Positive01% of infection8.3No. Positive02% of infection16.7Total infection (%)06 (50)	No. Examined 12 14 No. Positive 03 04 % of infection 25 28.6 No. Positive - 04 % of infection - 28.6 No. Positive 01 02 % of infection 8.3 14.3 No. Positive 02 - % of infection 16.7 - Total infection (%) 06 (50) 10 (71.5)	Two Three Four No. Examined 12 14 12 No. Positive 03 04 03 % of infection 25 28.6 25 No. Positive - 04 02 % of infection - 28.6 16.7 No. Positive 01 02 01 % of infection 8.3 14.3 8.3 No. Positive 02 - 03 % of infection 8.3 14.3 25 % of infection 16.7 - 25	Two Three Four Five No. Examined 12 14 12 15 No. Positive 03 04 03 03 % of infection 25 28.6 25 20 No. Positive - 04 02 01 % of infection - 28.6 16.7 6.7 No. Positive 01 02 01 06 % of infection 8.3 14.3 8.3 40 No. Positive 02 - 03 - % of infection 16.7 - 25 - Total infection (%) 06 (50) 10 (71.5) 9 (75) 10(66.7)	Two Three Four Five Six No. Examined 12 14 12 15 03 No. Positive 03 04 03 03 02 % of infection 25 28.6 25 20 66.7 No. Positive - 04 02 01 - % of infection - 28.6 16.7 6.7 - No. Positive 01 02 01 06 - % of infection 8.3 14.3 8.3 40 - No. Positive 02 - 03 - - % of infection 16.7 - 25 - - % of infection 16.7 - 25 - - Total infection (%) 06 (50) 10 (71.5) 9 (75) 10(66.7) 02(66.7)	Two Three Four Five Six Seven No. Examined 12 14 12 15 03 04 No. Positive 03 04 03 03 02 - % of infection 25 28.6 25 20 66.7 - No. Positive - 04 02 01 - - % of infection - 28.6 16.7 6.7 - - No. Positive 01 02 01 06 - - % of infection 8.3 14.3 8.3 40 - - % of infection 16.7 - 25 - - - % of infection 16.7 - 25 - - - % of infection 16.7 - 25 - - - % of infection 16.7 - 25 - - - Total